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## Research article

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Full-Text HTML I PDFNyanga pottery and the Manyika ethnohistory: towards a decolonised archaeology of the Nyanga agricultural complex
Robert T. Nyamushosho, Njabulo Chipangura, Takudzwa B. Pasipanodya, Foreman Bandama, Shadreck Chirikure, Munyaradzi Manyanga
e06609
Full-Text HTML I PDFEffects of phosphorus and sulfur on yield and nutrient uptake of wheat (Triticum aestivum L.) on Vertisols, North Central, Ethiopia
Shawl Assefa, Wassie Haile, Wondwosen Tena
e06614
Full-Text HTML I PDF
onservation agriculture-related practices contribute to maize (Zea mays L.) yield and soil improvement Central Malawi

Harrington Nyirenda, Victoria Balaka
e06636
Full-Text HTML I PDF
$\square$ Breaking out: the turning point in learning using mobile technology
Julia Bello-Bravo, Ian Brooks, Anne Namatsi Lutomia, Jeremy Bohonos, John Medendorp, Barry Pittendrigh
e06595
Full-Text HTML I PDF


Impact of intrapersonal and interpersonal emotional intelligence and self-directed learning on academic performance among pre-university science students
Emmanuel Nkemakolam Okwuduba, Kingsley Chinaza Nwosu, Ebele Chinelo Okigbo, Naomi Nkiru Samuel, Chinwe Achugbu e06611

Full-Text HTML I PDFEfficient removal of lead and arsenic using macromolecule-carbonized rice husks
Zeinab Babazad, Fariborz Kaveh, Mehdi Ebadi, Ramin Zafar Mehrabian, Mohammad Habibi Juibari
e06631
Full-Text HTML I PDF


Occurrence and geochemical significance of fluorene and alkylfluorenes in crude oils and source rock extracts from Niger Delta basin, Nigeria
Abiodun B. Ogbesejana, Oluwasesan M. Bello, Oluwole Joshua Okunola
e06616
Full-Text HTML I PDF
$\square$ Prediction of potential inhibitors of SARS-CoV-2 using 3D-QSAR, molecular docking modeling and
$\qquad$ DMET properties

Ayoub Khaldan, Soukaina Bouamrane, Fatima En-Nahli, Reda El-mernissi, Khalil El khatabi, Rachid Hmamouchi, Hamid Maghat, Mohammed Aziz Ajana, Abdelouahid Sbai, Mohammed Bouachrine, Tahar Lakhlifi e06603

Full-Text HTML I PDFPrevalence and associated factors of intimate partner violence (IPV) against women in Bangladesh amid COVID-19 pandemic

Istihak Rayhan, Khaleda Akter
e06619
Full-Text HTML I PDFThe real value of words: how target language linguistic modelling of foreign language teaching content shapes students' professional identity

Alexander Y. Bagiyan, Tatyana A. Shiryaeva, Elena V. Tikhonova, Natalia M. Mekeko e06581

Full-Text HTML I PDFEnvironmental correlation and epidemiologic analysis of COVID-19 pandemic in ten regions in five continents

Nadim Sharif, Mithun Kumar Sarkar, Shamsun Nahar Ahmed, Rabeya Nahar Ferdous, Nasir Uddin Nobel, Anowar Khasru Parvez, Ali Azam Talukder, Shuvra Kanti Dey
e06576
Full-Text HTML I PDFImpacts of the American Joint Committee on Cancer (AJCC) $8^{\text {th }}$ edition tumor, node, metastasis (TNM) staging system on outcomes of differentiated thyroid cancer in Thai patients

Yotsapon Thewjitcharoen, Waralee Chatchomchuan, Krittadhee Karndumri, Sriurai Porramatikul, Sirinate Krittiyawong, Ekgaluck ‘^’anothayaroj, Siriwan Butadej, Soontaree Nakasatien, Veekij Veerasomboonsin, Auchai Kanchanapituk, Rajata Rajatanavin, Thep mathongkam

16624

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Assia Aboubakar Mahamat, Ifeyinwa Ijeoma Obianyo, Blasuis Ngayakamo, Numfor Linda Bih, Olugbenga Ayeni, Salifu T.
Azeko, Holmer Savastano Jr.
e06597
Full-Text HTML I PDFHealth care provider's risk perception, and preparedness towards COVID-19 pandemic in North Central Ethiopia, 2020
Binyam Minuye Birihane, Wubet Alebachew Bayih, Yohannes Tesfahun, Tigabu Munye, Abebaw Yeshambel Alemu, Demeke Mesfin Belay
e06610
Full-Text HTML I PDFStudy of artifacts in thermodynamic and structural properties of Li-Mg alloy in liquid state using linear and exponential models
R.K. Gohivar, S.K. Yadav, R.P. Koirala, D. Adhikari e06613

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$\square$ Assessment of global warming in AI Buraimi, sultanate of Oman based on statistical analysis of NASA POWER data over 39 years, and testing the reliability of NASA POWER against meteorological measurements

Osama A. Marzouk
e06625
Full-Text HTML I PDF
:二 utritional compositions of two edible insects: Oryctes rhinoceros larva and Zonocerus variegatus neka Godwin Anaduaka, Nene Orizu Uchendu, Dionysius Obinna Osuji, Lorreta Nwakaego Ene, Ogechukwu Peace Amoke
e06531
Full-Text HTML I PDF


Magnetite nanoparticles grafted with murexide-terminated polyamidoamine dendrimers for removal of lead (II) from aqueous solution: synthesis, characterization, adsorption and antimicrobial activity studies Selma Ekinci, Zülfiye İlter, Selami Ercan, Ercan Çınar, Reşit Çakmak e06600

Full-Text HTML I PDFScientific justifications for the political decision-making on environmental remediation carried out after the Fukushima nuclear accident

Maria R.H. Takeuchi, Tatsuya Hasegawa, Susie M.L. Hardie, Linda E. McKinley, Gian Powell B. Marquez, Keiichi N. Ishihara e06588

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Theingi Win Myat, Nway Nway Thin Aung, Hlaing Myat Thu, Aye Aye, Nyo Nyo Win, Maung Maung Lwin, Htin Lin, Nang Sarm Hom, Kyaw Swar Lin, Moh Moh Htun e06601

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Francisco Javier Sáez-Fernández, Andrés J. Picazo-Tadeo, Ignacio Jiménez-Hernández e06524

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ənevieve Beaulieu-Pelletier, Marc-André Bouchard, Frederick L. Philippe
e06599
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Ishrat Jahan, Md. Rabiul Islam, Md. Reazul Islam, Rubaiya Ali, S.M. Matiur Rahman, Zabun Nahar, Abul Hasnat, Md. Saiful Islam e06621

Full-Text HTML I PDFHighly efficient synthesis of biodiesel catalyzed by a cellulose@hematite-zirconia nanocomposite Helmiyati Helmiyati, Yuni Budiman, Gusma Harfiana Abbas, Fitriyah Wulan Dini, Munawar Khalil e06622

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Cornelia Sindermann, Helena Sophia Schmitt, Dmitri Rozgonjuk, Jon D. Elhai, Christian Montag e06503

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The effect of direct and extended contact on attitudes towards social robots Marina Sarda Gou, Thomas L. Webb, Tony Prescott

16418
ll-Text HTML I PDF

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V. Kimpouni, J.D.D. Nzila, N. Watha-Ndoudy, M.I. Madzella-Mbiemo, S. Yallo Mouhamed, J.-P. Kampe
e06579
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M. Ángeles Jurado-Caraballo, Mercedes Rodríguez-Fernández
e06584
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e06583
Full-Text HTML I PDFEducational needs on safe motherhood from the perspective of suburban women: A qualitative study
Zohreh Mahmoodi, Mohsen Arabi, Kourosh Kabir, Mansoureh Yazdkhasti, Mahnaz Akbari Kamrani, Zahra Mehdizadeh Tourzani, Sara Esmaelzadeh
e06582
Full-Text HTML I PDFDielectric relaxation model of human blood as a superposition of Debye functions with relaxation times following a Modified-Weibull distribution
Charandeep Singh Sodhi, Luan Carlos de Sena Monteiro Ozelim, Pushpa Narayan Rathie e06606

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Manuel Gonzalez-Igual, Teresa Corzo Santamaria, Antonio Rua Vieites
e06495
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Emigdio Larios-Gómez, Laura Fischer, Mónica Peñalosa, Mayra Ortega-Vivanco
e06468
Full-Text HTML I PDF
$\square$
Estimating canopy nitrogen concentration of sugarcane crop using in situ spectroscopy
Aldemar Reyes-Trujillo, Martha C. Daza-Torres, Carlos A. Galindez-Jamioy, Esteban E. Rosero-García, Fernando MuñozArboleda, Efrain Solarte-Rodriguez
e06566
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K. Fabrice Kapiamba, Merveille Kimpiab
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Pei-Pei Sun, Ching-Shu Lai, Chung-Jung Hung, Periyathambi Dhaiveegan, Mei-Ling Tsai, Chun-Lun Chiu, Jim-Min Fang e06577

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Aloke Purkait, Ayan Mukherjee, Dipak Kumar Hazra, Kusal Roy, Pabitra Kumar Biswas, Ramen Kumar Kole e06557

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Higemengist Astatkie, Argaw Ambelu, Embialle Mengistie Beyene e06385

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e06575
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Alex Okello, Brian Owino Owuor, Jane Namukobe, Denis Okello, Julius Mwabora
e06571
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innmoy Biswas, Sadia Sarmin Soma, Md. Fazle Rohani, Md. Hamidur Rahman, Abul Bashar, Md. Sazzad Hossain 16587

Full-Text HTML I PDFMutational analysis of structural proteins of SARS-CoV-2
Shweta Jakhmola, Omkar Indari, Dharmendra Kashyap, Nidhi Varshney, Ayan Das, Elangovan Manivannan, Hem Chandra Jha e06572

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Dewi Setyaningsih, Yosua Agung Santoso, Yustina Sri Hartini, Yosi Bayu Murti, Wouter L.J. Hinrichs, Christine Patramurti e06541

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ıahputra Wibowo, Sri Widyarti, Akhmad Sabarudin, Djoko Wahono Soeatmadji, Sutiman Bambang Sumitro
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$\square$ Cloning, heterologous expression, and characterization of a novel thioesterase from natural sample Suharti, Gita Mahardika, Raissa, Laksmi Dewi, Heni Yohandini, Made Puspasari Widhiastuty, Raden Aditya Wibawa Sakti, Setyanto Tri Wahyudi, Akhmaloka
e06542
Full-Text HTML I PDF
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Poournima Patil, Suresh Killedar
e06526
Full-Text HTML I PDFThe new identity of Indonesian Islamic boarding schools in the "new normal": the education leadership response to COVID-19
Yusuf Hanafi, Ahmad Taufiq, Muhammad Saefi, M. Alifudin Ikhsan, Tsania Nur Diyana, Titis Thoriquttyas, Faris Khoirul Anam e06549

Full-Text HTML I PDFFactors associated with age of mother at first birth in Albania: application of quantile regression model Ashis Talukder, Zahidul Islam Khan, Fatheha Khatun, Shafia Tahmida e06547

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inu, Akbar Ali, Shakeel Ahmed
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Maximus Monaheng Sefotho
e06540
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Azhar Dyusupova, Raida Faizova, Oksana Yurkovskaya, Tatiana Belyaeva, Tatiana Terekhova, Amina Khismetova, Antonio SarriaSantamera, Dmitry Bokov, Alexandr Ivankov, Natalya Glushkova
e06561
Full-Text HTML I PDFA novel quadband ultra miniaturized planar antenna with metallic vias and defected ground structure for portable devices
Pierre Moukala Mpele, Franck Moukanda Mbango, Dominic B.O. Konditi, Fabien Ndagijimana e06373

Full-Text HTML I PDFEvaluating the shelf-life of pasteurized milk in Oman
M. Al-Farsi, I. Al-Gharibi, A. Al-Abri, A. Al-Humaimi, F. Al-Nabhani, H. Al-Hashmi, K. Al-Sarmi, S. Al-Shibli e06555

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16569
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Berenice Rojo-Garibaldi, Costanza Rangoni, Diego L. González, Julyan H.E. Cartwright
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Tareq Manzoor, K. Nazar, S. Iqbal, Habib Ullah Manzoor
e06567
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Yukio Fujiwara, Ryoko Maeda, Hidenori Takeshita, Yoshihiro Komohara
e06551
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Nayana Damenu Bandara Jayasundara, Palitha Arampath
e06560
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Sisay Kidanu, Ferdu Azerefegne, Esayas Mendesil
e06546
Full-Text HTML I PDF
an a monologue-style ECA more effectively motivate eHealth users in initial distress than textual Jidance?

Mark R. Scholten, Saskia M. Kelders, Julia E.W.C. Van Gemert-Pijnen
e06509
Full-Text HTML I PDFL-Citrulline ameliorates the attenuation of acetylcholine-induced vasodilation of retinal arterioles in diabetic rats

Asami Mori, Toshiaki Takei, Namiko Suzuki, Kenji Sakamoto, Masahiko Morita, Satoshi Nakagawa, Tsutomu Nakahara, Kunio Ishii e06532

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$\square$ What factors influence access to and the level of participation in high value mango markets by smallholder farmers in Ghana?

Rexford Akrong, Stephen G. Mbogoh, Patrick Irungu
e06543
Full-Text HTML I PDFReligiosity and stigma toward patients with mental illness among undergraduate university students Ahlam AI-Natour, Sawsan Abuhammad, Hanan Al-Modallal
e06565
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Mariasole Cervini, Antonello Frustace, Guillermo Duserm Garrido, Gabriele Rocchetti, Gianluca Giuberti e06562

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oes worriedness among the rural adults promote COVID-19 related awareness in Bangladesh? uhammad Mahmudul Hasan, Ashis Talukder, Muhammad Khairul Alam, Muhammad Kausar Hossain, Asikunnaby
e06556
Full-Text HTML I PDFSynthesis, structural characterization, and DFT studies of anti-cancer drug N-(2-Aminophenyl)-2-(4bromophenoxy)acetamide
S.N. Chandana, Fares Hezam Al-Ostoot, Yasser Hussein Eissa Mohammed, Tareq N. Al-Ramadneh, P. Akhileshwari, Shaukath Ara Khanum, M.A. Sridhar, B.N. Lakshminarayana
e06464
Full-Text HTML I PDF
$\square$
Full exclusion during COVID-19: Saudi Deaf education is an example
Abdullah Madhesh
e06536
Full-Text HTML I PDFSowing methods and seeding rates effects on yield and yield components of Tef(Eragrostis tef [Zucc.] Trotter) at Adet, North West Ethiopia
Yechale Mengie, Alemayehu Assefa, Abaynew Jemal Jenber e06519

Full-Text HTML I PDF
$\square$ Growth characteristics of human bone marrow mesenchymal stromal cells at cultivation on synthetic polyelectrolyte nanofilms in vitro
Lyudmila M. Mezhevikina, Dmitriy A. Reshetnikov, Maria G. Fomkina, Nurbol O. Appazov, Saltanat Zh. Ibadullayeva, Evgeniy E. Fesenko
e06517
Full-Text HTML I PDF
¿= oil fertility management among smallholder farmers in Mount Kenya East region

Amos W. Wawire, Ádám Csorba, József A. Tóth, Erika Michéli, Márk Szalai, Evans Mutuma, Eszter Kovács e06488

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Jonah Chukwudi Umeuzuegbu, Stanley Okiy, Chidozie Chukwuemeka Nwobi-Okoye, Okechukwu Dominic Onukwuli e06516

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Faisal Alshawabkeh, Mohmmad Husien Almajali
e06471

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Rony Escobar-Yonoff, Daniel Maestre-Cambronel, Sebastián Charry, Adriana Rincón-Montenegro, Ivan Portnoy e06506

Full-Text HTML I PDF
$\square$ Biochemical and antioxidant properties of cream and orange-fleshed sweet potato
$\qquad$ əbecca Olajumoke Oloniyo, Olufunmilayo Sade Omoba, Olugbenga Olufemi Awolu 16533

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Clemente Rodríguez-Sabiote, José Álvarez-Rodríguez, Daniel Álvarez-Ferrandiz, Félix Zurita-Ortega
e06469

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Nathalia P. Scioscia, Patricia E. Pensel, Guillermo M. Denegri, María Celina Elissondo e06496

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Rita Rodrigues, Ana P. Chung, Martin S. Mortensen, Maria H. Fernandes, António B. Monteiro, Rowney Furfuro, Cátia C. Silva, Maria C. Manso, Søren J. Sørensen, Paula V. Morais
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M. Teil, A. Regazzi, B. Harthong, P.J.J. Dumont, D. Imbault, J.-L. Putaux, R. Peyroux e06482

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Kihinetu Gelaye Wudineh, Fentahun Alemnew Chekole, Azimeraw Arega Tesfu
e06528
Full-Text HTML I PDFWillingness to join and pay for community-based health insurance and associated determinants among urban households of Cameroon: case of Douala and Yaounde
jsine Wafo Cheno, William Tchabo, Jonathan Tchamy
16507

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Aryan Zahergivar, Madison Kocher, Jeffrey Waltz, Ismail Kabakus, Jordan Chamberlin, Selcuk Akkaya, Ali M. Agha, U.Joseph Schoepf, Jeremy R. Burt
e06386
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e06452

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Omar F. Khabour, Salwa F.M. Hassanein
e06538
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he exacerbation of violence against women as a form of discrimination in the period of the COVID-19
$\qquad$ andemic

Paula Andrea Valencia Londoño, Martha Elisa Nateras González, Constanza Bruno Solera, Phoenix Storm Paz e06491

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Lisa Brauer Oliveira, Mauro Geller, Karin Soares Cunha, Alessandra Santos, Allan Bernacchi, Allan E. Rubenstein, Sanyu Takirambudde, Spyros Mezitis, Carolina de Almeida Ito Brum, Luiz Guilherme Darrigo Jr., Marcia Gonçalves Ribeiro e06518

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e06497
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Omonike A. Olaleye, Manvir Kaur, Collins Onyenaka, Tolulope Adebusuyi
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16511
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e06522
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e06530
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Yoshikatsu Akiyama
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:二
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Quynh Hoa Tran, Van Gio Nguyen, Cong Manh Tran, Minh Nam Nguyen
e06463
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16483
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:二 16466

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16438
:二
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:二

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:二
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Khaled Alkethiri, Tariq Almtroudi, Abdullah bin Jurays, Faisal Abanumay, Mohammed Aldammas, Meshaal AlKhodheer, Muhammad Iqbal, Syed Shahid Habib, Shahid Bashir e06358

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$\square$ Drought, hunger and coping mechanisms among rural household in Southeast Ethiopia
Betemariam Gebre, Habtamu Yesigat Ayenew, Sibhatu Biadgilign
e06355
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Amare Bihon, Solomon Zinabu, Yimer Muktar, Ayalew Assefa
e06325
Full-Text HTML I PDFCFD and statistical approach to optimize the average air velocity and air volume fraction in an inertparticles spouted-bed reactor (IPSBR) system
A. Mohammad, A.A.H.I. Mourad, A.H. Al-Marzouqi, M.H. El-Naas, B. Van der Bruggen, M. Al-Marzouqi, F. Alnaimat, M. Suleiman, M. Al Musharfy e06369

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Sameh A. Rizk, Maher A. El-Hashash, Amr A. Youssef, Abdelfattah T. Elgendy
e06220
Ill-Text HTML I PDF

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e06341
Full-Text HTML I PDF
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e06360
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Touria Ouslimane, Lhoussayne Et-taya, Lahoucine Elmaimouni, Abdellah Benami
e06379
Full-Text HTML I PDFCytoplasmic genome of Indian potato varieties and breeding lines vis a vis prospects in potato breeding Salej Sood, Ashwani Kumar, Baljeet Singh, Sundaresha S, Vinay Bhardwaj e06365

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## Corrigendum

Corrigendum to "Modeling, simulation and optimization of solid fuel bread ovens commonly used in developing countries" [Heliyon 7(2), (February 2021) Article e06184]C.F. Kouemou Hatou, G. Tchuen, P. Woafo

16585
ill-Text HTML I PDF

Corrigendum to "Investigating the effect of meditation on spiritual wellbeing of Type-2 diabetic amputees: A clinical trial study" [Heliyon 6 (11) (November 2020) e05567]

Ali Heydari Movahed, Fakhri Sabouhi, Reza Mohammadpourhodki, Sepideh Mahdavi, Sima Goudarzian, Malihe Amerian, Mona Mohtashami, Mansoure Kheiri, Malihe Imeni
e06508
Full-Text HTML I PDF
$\square$ Corrigendum to "Towards identifying enablers and inhibitors to on-farm entrepreneurship: Evidence from smallholders in KwaZulu-Natal, South Africa" [Heliyon 7 (1) (January 2021) Article e05660]
Edilegnaw Wale, Unity Chipfupa, Nolwazi Hadebe
e06376
Full-Text HTML \| PDF

## Review article

Mobile health applications for disease screening and treatment support in low-and middle-income countries: A narrative reviewErnest Osei, Tivani P. Mashamba-Thompson
e06639
Full-Text HTML I PDFDoes social capital improve farm productivity and food security? Evidence from cocoa-based farming households in Southwestern Nigeria
A.D. Kehinde, R. Adeyemo, A.A. Ogundeji
e06592
Full-Text HTML I PDF
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ioremediation and pharmacological applications of gold nanoparticles synthesized from plant materia

Sunday Adewale Akintelu, Bo Yao, Aderonke Similoluwa Folorunso
e06591
Full-Text HTML I PDFChoices of chromatographic methods as stability indicating assays for pharmaceutical products: A review

Yik-Ling Chew, Mei-Ann Khor, Yau-Yan Lim
e06553
Full-Text HTML I PDFHeat stress on cattle embryo: gene regulation and adaptation
Juan Sebastian Naranjo-Gómez, Heinner Fabián Uribe-García, María Paula Herrera-Sánchez, Kelly Johanna Lozano-Villegas, Roy Rodríguez-Hernández, lang Schroniltgen Rondón-Barragán
e06570
Full-Text HTML I PDFEnvironmental health situation in Nigeria: current status and future needs
Hyellai Titus Pona, Duan Xiaoli, Olusola O. Ayantobo, Narh Daniel Tetteh
e06330
Full-Text HTML I PDFFuture call for policy making to speed up interdisciplinarity between natural and social sciences and humanities in countries such as India
Kabita Das, Biswaranjan Paital
e06484
Full-Text HTML I PDF
otential of long non-coding RNAs as a therapeutic target and molecular markers in glioblastoma athogenesis

Rishabh Chaudhary
e06502
Full-Text HTML I PDFNanoparticles as antimicrobial and antiviral agents: A literature-based perspective study
Shabnam Sharmin, Md. Mizanur Rahaman, Chandan Sarkar, Olubunmi Atolani, Mohammad Torequl Islam, Oluyomi Stephen Adeyemi
e06456
Full-Text HTML I PDF


Dose-response meta-analysis of arsenic exposure in drinking water and hypertension
Afsaneh Amiri, Yaser Mokhayeri, Rasool Mohammadi, Mohammad Amin Karami, Mansour Ghaderpoori, Bahram Kamarehie, Ali Jafari
e06409
Full-Text HTML I PDF


Application and control of flexible alternating current transmission system devices for voltage stability enhancement of renewable-integrated power grid: A comprehensive review
Bukola Babatunde Adetokun, Christopher Maina Muriithi
e06461
Full-Text HTML I PDFPercutaneous microwave ablation applications for liver tumors: recommendations for COVID-19 patients
Pooya Afaghi, Michael Anthony Lapolla, Khashayar Ghandi e06454

Full-Text HTML I PDF

Liver regeneration observed across the different classes of vertebrates from an evolutionary perspective
Blanca Delgado-Coello
e06449
Full-Text HTML I PDFNatural language processing for urban research: A systematic review
Meng Cai
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$\square$
Vermiwash: An agent of disease and pest control in soil, a review
Kasahun Gudeta, J.M. Julka, Arun Kumar, Ankeet Bhagat, Amita Kumari
e06434
Full-Text HTML I PDFMultifaceted roles of Toll-like receptors in acute kidney injury
Rakhshinda Habib
e06441
Full-Text HTML I PDFDrivers of anthropogenic air emissions in Nigeria - A review
Oyetunji B. Okedere, Francis B. Elehinafe, Seun Oyelami, Augustine O. Ayeni
e06398
Full-Text HTML I PDF
$\square$ Failed induction of labor and its associated factors in Ethiopia: A systematic review and meta-analysis
$\qquad$ נenezer Melkie, Dagne Addisu, Maru Mekie, Enyew Dagnew 16415

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Ida Hamidah, Roer Eka Pawinanto, Budi Mulyanti, Jumril Yunas
e06406
Full-Text HTML I PDFImpact of SARS-CoV-2 on the clinical outcomes and placental pathology of pregnant women and their infants: A systematic review
Irina Oltean, Jason Tran, Sarah Lawrence, Brittany Ann Ruschkowski, Na Zeng, Cameron Bardwell, Youssef Nasr, Joseph de Nanassy, Dina El Demellawy
e06393
Full-Text HTML I PDFA review of augmented reality systems and their effects on mental workload and task performance Nor Farzana Syaza Jeffri, Dayang Rohaya Awang Rambli
e06277
Full-Text HTML I PDFRecent progress in surface plasmon resonance based sensors: A comprehensive review Vasimalla Yesudasu, Himansu Shekhar Pradhan, Rahul Jasvanthbhai Pandya e06321

Full-Text HTML I PDFMarine endophytic fungal metabolites: A whole new world of pharmaceutical therapy exploration Esraa Ahmed Mohamed El-Bondkly, Alaa Ahmed Mohamed El-Bondkly, Aya Ahmed Mohamed El-Bondkly e06362
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Gyan Datta Tripathi, Zoya Javed, Mansi Mishra, Vinayak Fasake, Kavya Dashora
e06150
Full-Text HTML I PDFBrain-to-brain communication: the possible role of brain electromagnetic fields (As a Potential Hypothesis)
Ehsan Hosseini
e06363
Full-Text HTML I PDFCorrigendum to "Blood hormones and torque teno virus in peripheral blood mononuclear cells" [Heliyon 6 (11) (2020) e05535]
Peik M.A. Brundin, Britt-Marie Landgren, Peter Fjällström, Anders F. Johansson, Ivan Nalvarte e06568

Full-Text HTML I PDFThe relationship of oleic acid/albumin molar ratio and clinical outcomes in leptospirosis
Caroline Azevedo Martins, Maria Conceição B dos Santos, Cassiano Felippe Gonçalves-de-Albuquerque, Hugo Caire Castro-FariaNeto, Mauro Velho Castro-Faria, Patricia Burth, Mauricio Younes-Ibrahim
e06420
Full-Text HTML I PDF
eport

A novel approach to teaching Hidden Markov Models to a diverse undergraduate population
Philip Heller, Pratyusha Pogaru
e06437
Full-Text HTML I PDF

## Erratum

Erratum to "Development of a novel polymerase spiral reaction (PSR) assay for rapid and visual detection of Clostridium perfringens in meat" [Heliyon 7 (1) (January 2021) Article e05941]A. Arun Prince Milton, Kasanchi M. Momin, Sandeep Ghatak, G. Bhuvana Priya, M. Angappan, Samir Das, K. Puro, R.K. Sanjukta, I. Shakuntala, A. Sen, B.K. Kandpal e06332

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Research article

# Isocratic high-performance liquid chromatography (HPLC) for simultaneous quantification of curcumin and piperine in a microparticle formulation containing Curcuma longa and Piper nigrum 

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## A R T I C L E I N F O

## Keywords:

Curcumin
Piperine
Extract
Solid dispersion
Reverse-phase
Validation
Optimization


#### Abstract

Poor bioavailability has been reported as a major challenge in the development of curcumin as a pharmaceutical agent. However, co-administration of curcumin with piperine has been shown to improve curcumin bioavailability. Therefore, to assure product control quality, an analytical method needs to be developed for the determination of curcumin and piperine content in a dosage form formulation. The objective of this study was to develop a simple isocratic reversed-phase HPLC (RP-HPLC) method to simultaneously quantify curcumin and piperine content in solid dispersion based microparticle formulation containing Curcuma longa and Piper nigrum extracts. The method was validated according to the International Council for Harmonization (ICH) guideline. Chromatographic separation of three curcuminoids and piperine could be achieved using acetonitrile-methanolwater of 65:5:35 \%, at a flow rate of $1 \mathrm{~mL} / \mathrm{min}$ and a wavelength of 353 nm for detection. Resolution (Rs) of 3.57 and 1.68 for piperine and curcumin, respectively, a theoretical plate number $(\mathrm{N})>8000$ and a tailing factor $(\mathrm{T})<$ 1.5 indicate a satisfactory separation of the compounds. The calibration curve was linear from $1.25-15 \mu \mathrm{~g} / \mathrm{mL}$ and $2.5-30 \mu \mathrm{~g} / \mathrm{mL}$ for piperine and curcumin, respectively, with the correlation coefficient of $>0.999$. The intra-day/ inter-day accuracy and precision demonstrated a recovery of 99.54-101.50\%/99.38-99.89\% and $100.78-102.51 \% / 101.15-102.47 \%$ with a Relative Standard Deviation (RSD) of 0.53-0.95\%/0.13-1.44 \% and $0.28-1.62 \% / 0.46-1.14 \%$ for piperine/curcumin. The limit of detection (LOD) were 0.27 and $0.42 \mu \mathrm{~g} / \mathrm{mL}$, for piperine and curcumin, which reveals an adequate sensitivity. A solid dispersion based microparticle formulation containing C. longa and P. nigrum extracts confirmed the validity of the developed method as a recovery of $91.14 \%$ and $99.14 \%$ for piperine and curcumin, respectively. In conclusion, all the tested parameters confirm the precision, accuracy, and reliability of the method for the simultaneous analysis of curcumin and piperine within a microparticle formulation containing C. longa and $P$. nigrum extracts.


## 1. Introduction

Curcumin, a yellow polyphenolic compound isolated from turmeric (Curcuma longa) rhizome, has been claimed as a therapeutic agent for numerous diseases due to its anti-oxidant and anti-inflammatory properties [1, 2, 3]. However, despite extensive in-vitro and in-vivo studies, a clear therapeutic effect of curcumin in clinical studies has not been found yet [4]. These disappointing results can be ascribed to the poor bioavailability of the compound after oral intake. There are two reasons for the poor absorption of curcumin; 1) It exhibits a very low aqueous
solubility of only $11 \mathrm{ng} / \mathrm{mL}$ [4, 5], and 2) It is rapidly metabolized [1]. Therefore, a combination of two strategies addressing poor solubility and rapid metabolism is required.

An often-applied strategy to improve the dissolution behavior of a poorly water-soluble drug is formulating it as a solid dispersion [6]. Furthermore, piperine (Figure 1d), an alkaloid compound of black pepper ( $P$. nigrum) extract has been identified as a bio-enhancer of various drugs, by stimulating gut amino acid transporters and inhibiting drug-metabolizing enzymes to improve drug absorption, bioavailability, and bio-efficacy [7]. Both strategies are thought to be useful to be applied

[^1]
(a)

(b)

(d)

Figure 1. Molecular structure of (a) curcumin; (b) demethoxycurcumin; (c) bis-demethoxycurcumin; (d) piperine.
to curcumin. Hence, we propose a formulation based on solid dispersion technology containing $C$. longa and $P$. nigrum extracts to improve the bioavailability of curcumin. In this study, we will focus on solid dispersion microparticles prepared by spray drying using polyvinylpyrrolidone K30 (PVP K30) as a carrier.

During pre-formulation, the quality control of the product needs to be addressed. Determining compounds in the final product is a part of quality assurance in production processes. Therefore, the amount of curcumin and piperine containing in the microparticle needs to be determined accurately.

Several analytical methods for the quantification of curcumin and piperine have been developed, however, the majority of these methods concern the quantification of curcumin and piperine individually $[8,9,10$, 11]. A few methods have been developed to simultaneously determine curcumin and piperine. A spectroscopic method combining with the application of Vierordt's equation was successfully applied to allow the determination of curcumin and piperine as a mixture of $C$. longa and $P$. nigrum extracts in dissolution medium [12], however, the method is not selective.

HPLC with UV-Vis detection is one of the most common techniques used in the analysis and quality control of several formulations containing curcumin or piperine and in biological samples. Moorthi et al. validated an isocratic HPLC method to allow curcumin and piperine quantification in a Eudragit based nanosuspension containing pure curcumin and piperine as a binary mixture [13]. Furthermore, Sethi et al. reported a selective and specific HPLC method for simultaneous estimation of curcumin and piperine in human plasma [14]. Also, a sensitive LC/MS method was developed to enable simultaneous quantification of curcumin and piperine for pharmacokinetic evaluation [15]. Although the HPLC methods have been reported to be useful to simultaneously analyze curcumin and piperine in various samples, the developed methods were applied to determine curcumin and piperine as pure compounds in the samples.

To our best knowledge, no HPLC analytical method has been reported for the simultaneous analysis of curcumin and piperine in extracts. It should be emphasized that commercially available curcumin originating from turmeric rhizome is usually composed of a mixture of naturally occurring curcuminoids, namely curcumin, demethoxycurcumin, and bisdemethoxycurcumin (Figure 1a, b, c). This mixture of compounds could easily interfere with chromatographic separation if not properly resolved. Therefore, the study aimed to develop a validated HPLC method to simultaneously quantify curcumin and piperine in solid dispersion-based microparticle formulation containing $C$. longa and $P$. nigrum extracts.

## 2. Materials and methods

### 2.1. Materials

Bis-demethoxycurcumin, demethoxycurcumin, curcumin, and piperine reference standards with a purity of $>98 \%$ were purchased from

Sigma-Aldrich (St. Louis, USA). C. longa extract of high curcuminoids content (97.56\%) was a kind gift from PT Phytochemindo Reksa, Bogor, Indonesia. $P$. nigrum was extracted and purified as described by Saha et al. [16]. Polyvinylpyrrolidone K30 (PVP K30) was kindly given by PT Konimex, Solo, Central Java, Indonesia. Ethanol, HPLC grade acetonitrile, HPLC grade methanol, and analytical grade phosphoric acid were purchased from Merck (Darmstadt, Germany). A PTFE filter with a pore size of $0.22 \mu \mathrm{~m}$ was obtained from Whatman. Milli-Q water was prepared in the laboratory.

### 2.2. Preparation of solid dispersion based microparticle formulation and characterization

A PVP-30 based solid dispersion containing $30 \mathrm{w} / \mathrm{w} \%$ C. longa extract and $10 \mathrm{w} / \mathrm{w} \% P$. nigrum extract was prepared by spray drying. In brief, 1800 mg C. longa extract and 600 mg P. nigrum extract were dissolved in 400 mL ethanol and 3600 mg of PVP K30 was dissolved in 50 mL ethanol. The two solutions were mixed for approximately 15 min using a magnetic stirrer.

The resulting solution was fed into a BUCHI B-290 mini spray dryer equipped with a B-295 dehumidifier via a two-way channel with a nozzle diameter of 0.7 mm . Drying was conducted at an inlet temperature of 105 ${ }^{\circ} \mathrm{C}$, aspiration of $100 \%$, feeding rate of $20 \%$, and an atomization pressure of $500 \mathrm{Ln} / \mathrm{h}$. The outlet temperature was observed at $60-65^{\circ} \mathrm{C}$. Micrographs of the microparticles were taken using a scanning electron microscope (Hitachi TM 3000, Japan).

### 2.3. Method development

### 2.3.1. Finding the detection wavelength

Individual methanolic solutions of curcumin $(15 \mu \mathrm{~g} / \mathrm{mL})$ and piperine (5 $\mu \mathrm{g} / \mathrm{mL}$ ) were scanned at the range of $315-450 \mathrm{~nm}$ using a UV-Vis spectrophotometer (Shimadzu, Japan). The detection wavelength for the simultaneous analysis of curcumin and piperine was selected based on the crossing point of the curcumin and piperine spectra of the spectral overlay.

### 2.3.2. HPLC instrument and conditions

HPLC analysis was carried out using a Shimadzu LC 2010HT HPLC system (Shimadzu, Kyoto, Japan) equipped with a serial dual plunger pump, an autosampler, and an SPD-20A/20AV series UV-Vis detectors. LC-solution software was used for peak integration. Chromatographic separation was achieved on a C18 column ( $250 \times 4.6 \mathrm{~mm}$, Eurospher 100 with $5 \mu \mathrm{~m}$ ) equipped with a pre-column (Knauer, Berlin, Germany). The mobile phase was filtered through a $0.22 \mu \mathrm{~m}$ filter and degassed by ultrasonication before use. The injection volume was $20 \mu \mathrm{~L}$. The column temperature was kept at $33^{\circ} \mathrm{C}$ during chromatographic operation.

### 2.3.3. Selection of mobile phase

The aim was to find a mobile phase composition, which provides an acceptable peak separation of multiple components i.e. curcuminoids
(bis-demethoxycurcumin, demethoxycurcumin, curcumin) and piperine with a short running time of less than 10 min . A sample consisting of a mixture of $10 \mu \mathrm{~g} / \mathrm{mL} C$. longa and $5 \mu \mathrm{~g} / \mathrm{mL}$ P. nigrum extracts in methanolic solution was used to evaluate mobile phases of different compositions.

Peak identification was carried out by running standard solutions at accordingly used mobile phase compositions. The mobile phase compositions were evaluated on the chromatographic separation parameters i.e. tailing factor (T), peak resolution (Rs), and theoretical plate number ( N ). The acceptance criteria are $\mathrm{T}<2$, Rs $>1.5$ and $\mathrm{N}>2000$ [17].

### 2.3.4. Preparation of stock and calibration solutions

Stocks of standard solution of curcumin and piperine at a final concentration of $1 \mathrm{mg} / \mathrm{mL}$ were prepared by dissolving an appropriate amount of curcumin and piperine in methanol as an individual solution. All solutions were prepared in light-protected vials to ensure the stability of curcumin and piperine [18].

### 2.4. Method validation

The developed method was validated as per Q2R1 ICH guidelines [19] including selectivity, system suitability, accuracy, precision, limit of detection (LOD), limit of quantification (LOQ), linearity, and robustness.

### 2.4.1. System suitability

Six replicate injections of the sample containing a mixture of curcumin and piperine at a concentration of 10 and $5 \mu \mathrm{~g} / \mathrm{mL}$ in the methanolic solution were simultaneously analysed. Retention time (Tr), peak area (AUC), Rs, T , and N were determined. Relative standard deviation (RSD) values of these parameters were calculated to evaluate the system suitability of the developed method. The suitability test was accepted when the RSD values of these parameters were less than $2 \%$.

### 2.4.2. Specificity and selectivity

Specificity refers to the ability of a method to distinguish between the compound(s) of interest and impurity present in the sample matrix [20]. For the specificity study, the chromatogram of blank samples originated from the formulation matrix (no drugs) was compared to the chromatogram of microparticle formulation containing $C$. longa and $P$. nigrum extracts. Selectivity is the ability of the method to clearly show separation between the different peaks. A value of Rs $>1.5$ is required to confirm the method to be sufficiently selective [20].

### 2.4.3. Preparation of calibration solutions of curcumin and piperine and linearity

Calibration solutions were prepared using solutions of curcumin and piperine in methanol at different concentrations. Stock solutions of curcumin and piperine were diluted with methanol and mixed to obtain samples with curcumin concentrations of $0.05-45 \mu \mathrm{~g} / \mathrm{mL}$ and piperine concentrations of $0.025-20 \mu \mathrm{~g} / \mathrm{mL}$ for. Linearity between concentration and peak area was tested using the regression line as provided by the least square analyses. The correlation coefficient (r) was used to judge linearity [21]. The concentration range with an $r>0.99$ was considered linear. The limit of detection (LOD) and the limit of quantification (LOQ) were determined using the standard deviation approach [20]. To do so, serially diluted concentrations of the calibration solutions were prepared at nearly targeted LOD or LOQ and analyzed using least square analysis. The standard deviation (SD) of the y-intercept and slope were determined with which the LOD and LOQ were calculated based on Eq. (1) and Eq. (2) below [22].
$L O D=3.3 \times S D /$ slope
$L O Q=10 \times S D /$ slope

### 2.4.4. Accuracy and precision

Accuracy refers to the closeness of the obtained value with that true value of the corresponding sample concentrations, while precision denotes the closeness agreement between independent test results obtained under the developed method. In this study, accuracy and precision were carried out using the assay sample spiking method. The assay sample was the microparticle formulation as described in section 2.2; dissolved in methanol. The methanolic sample solutions at the concentration of $9.9 \mu \mathrm{~g} /$ mL (sample weight/volume) were spiked with curcumin and piperine stock solutions to result in a mixture of final addition concentrations of 2.50; $15.00 ; 30.00$ and $1.25 ; 7.50 ; 15.00 \mu \mathrm{~g} / \mathrm{m}$ for curcumin and piperine.

Accuracy was evaluated based on the recovery percentage, and precision was studied using repeatability assay based on the RSD values of three different sample concentrations. Accuracy and precision were conducted on one-day analysis (intra-day) and three consecutive days (inter-day) of freshly prepared samples. The analysis was done in triplicate for each concentration level. The degree of accuracy and precision were judged according to the Association of Official Analytical Chemists requirement [21].

### 2.4.5. Robustness

The robustness of a developed analytical method refers to its ability to remain unaffected by deliberate changes of the chromatographic conditions which indicates its reliability during normal usage. Peaks retention time and AUC values of curcumin and piperine resulted from the changed parameters of mobile phase composition, flow rates, and detection wavelengths were analyzed for the RSD values. The RSD value of less than $2 \%$ is an indication of a robust method.

### 2.4.6. Assay of curcumin and piperine in the microparticle formulation

A 200 mg of the microparticle formulation was accurately weighed and dissolved in 50 mL of methanol in a volumetric flask. The solution was subjected to ultrasonication for 5 min . The solution was diluted 100 times with methanol to achieve a final concentration of $40 \mu \mathrm{~g} / \mathrm{mL}$ and then analyzed by HPLC. The content of curcumin and piperine was calculated based on the calibration curve constructed as described in section 2.4.3.

### 2.5. Data analysis

A Least Square Analysis supported by Excel Microsoft INC USA was used to analyze the linearity of the calibration curve. Mean and standard deviation was calculated using Excel Software (Microsoft Inc, USA).

## 3. Results and discussion

### 3.1. Preparation of solid dispersion based microparticle formulation and characterization

Microparticles of PVP-30 based solid dispersion containing curcuminoids of $C$. longa and piperine of $P$. nigrum extracts could be successfully prepared by spray drying as can be seen in the scanning electron microscopic images (see example in Figure 2).

### 3.2. Finding the detection wavelength

The spectral scanning of a single methanolic solution of curcumin and piperine is shown in Figure 3. As shown in Figure 3, the methanolic solution of curcumin ( $15 \mu \mathrm{~g} / \mathrm{mL}$ ) absorbed wavelength at the maximum value of 425 nm , while piperine dissolved in methanol at the concentration of $5 \mu \mathrm{~g} / \mathrm{mL}$ showed a maximum absorption at the wavelength of 342 nm . Both compounds exhibited an overlapping absorption area at around 315-360 nm, where curcumin and piperine can be simultaneously detected.


Figure 2. Scanning electron microscopic image of the spray dried PVP K30 based solid dispersion containing curcuminoids and piperine.


Figure 3. The overlay spectra of curcumin $(15 \mu \mathrm{~g} / \mathrm{ml})$ and piperine $(5 \mu \mathrm{~g} / \mathrm{ml})$ in methanolic solution.

As can be seen in Figure 3, the two spectra cross each other at 353 nm . Therefore, this wavelength was used for the simultaneous detection of curcumin and piperine in this study.

### 3.3. Selection of mobile phase

### 3.3.1. The use of methanol and acetonitrile

Simultaneous analysis of a mixture contains weak acids and weak basics drugs on an RP HPLC is always challenging [23]. The target analytes in this study are curcumin (weak acid) and piperine (weak basic) which are present as co-mixture with other curcuminoids compounds (bis-demethoxycurcumin, demethoxycurcumin) as the active components in the microparticle formulation. The combinations methanol-water and acetonitrile-water are the most commonly used solvent mixtures in RP HPLC analysis. However, curcuminoids are poorly resolved in methanol-water mixtures at a volume ratio of $50: 50$ and $60: 40$ with calculated polarity index of 7.55 and 7.06. Methanol was therefore replaced by acetonitrile. Although with an acetonitrile-water volume ratio of $60: 40$ (polarity index of 7.48) a sharp peak was obtained, all three curcuminoids were co-eluted at a retention time of 12.54 min .

### 3.3.2. The effect of $0.1 \%$ phosphoric acid as pH modifier

A study by Espinosa et al. revealed that the retention time of acidic and basic compounds is strongly dependent on the pH of the mobile phase [24]. The addition of acid into the mobile phase can improve peak separation and reduce peak tailing by modification of interaction between the target analyte and the stationary phase of the HPLC column [25, 26].

Therefore, a mobile phase composition as reported by Sethi et al., which was used to simultaneously analyze curcumin and piperine in the plasma sample [14], with slight modifications was evaluated. The mobile phase consisted of acetonitrile-methanol-water-phosphoric acid $0.1 \%$ at a volume ratio of 20:30:47.5:2.5 ( pH 4.0 ) and was delivered at $1 \mathrm{~mL} / \mathrm{min}$ in an isocratic mode. However, under these conditions, the three curcuminoids compounds were co-eluted at a retention time of 11.80 min , whereas no piperine peak appeared within a running time of 15 min , while a maximum running time of 10 min was aimed for. To evaluate the behavior of the weakly basic compound (piperine) at a lower pH , the composition of the mobile phase was adjusted to acetonitrile-methanol-phosphoric acid $0.1 \%$ at a volume ratio of 20:30:50 ( pH 3.0 ) and a calculated polarity index of 7.79 . However, a peak in the chromatogram that can be assigned to piperine remained absent during a run time of 15 min . Remarkably, when the mobile phase polarity was slightly modified by replacing the methanol with acetonitrile to result in a calculated polarity index of 8.00 of the mobile phase compositions of acetonitrile-water-phosphoric acid $0.1 \%$ at a volume of 50:47.5:2.5 ( pH 4.0 ), piperine was eluted already at 0.516 min .

LoBrutto et al. reported the effect of pH on the retention time of small basic compounds, aniline, and pyridine on an isocratic HPLC mode. At the pH range of 1.3-8.6 of the mobile phase with a composition of acetonitrilewater at a volume ratio of $10: 90$, it was reported that as the pH of the mobile phase decreased, the retention time of the targeted compounds increased due to counterion interactions resulted from acidic mobile

Table 1. Mobile phase compositions and the observed peak characteristics.

| Code | ACN | MeOH | water | Curcuminoids |  |  |  |  |  |  |  |  | Piperine |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  |  |  |  | BDMC |  |  | DMC |  |  | C |  |  |  |  |  |
|  |  |  |  | Tr | Rs | T | Tr | Rs | T | Tr | Rs | T | Tr | Rs | T |
| 1 | 30 | 25 | 45 | - | - | - | 6.30 | 4.66 | 0 | 6.63 | 0.76 | 0 | - | - | - |
| 2 | 40 | 30 | 30 | 8.09 | 1.55 | 0 | 8.25 | 1.08 | 0 | 8.99 | 1.14 | 0 | 10.72 | 5.34 | 1.17 |
| 3 | 30 | 50 | 20 | 4.26 | 5.61 | 1.11 | 4.86 | 0 | 0 | 5.35 | 0 | 0.91 | 8.77 | 3.66 | 1.11 |
| 4 | 60 | 10 | 30 | 5.03 | 6.11 | 1.06 | 5.38 | 1.56 | 1.20 | 5.78 | 1.64 | 1.18 | 7.84 | 3.22 | 1.08 |
| 5 | 65 | 5 | 30 | 4.82 | 4.74 | 0.95 | 5.18 | 1.48 | 1.12 | 5.58 | 1.68 | 1.16 | 7.91 | 3.57 | 1.06 |

Note:

1. $\mathrm{ACN}=$ acetonitrile; $\mathrm{MeOH}=$ methanol.
2. $\mathrm{BDMC}=$ bis-demethoxycurcumin; $\mathrm{DMC}=$ demethoxycurcumin; $\mathrm{C}=$ curcumin.
3. $\mathrm{Tr}=$ retention time; $\mathrm{Rs}=$ resolution; $\mathrm{T}=$ tailing factor.
4.     - = not detected.


Figure 4. Chromatographic separation of bis-demethoxycurcumin (BDMC), demethoxycurcumin (DMC), curcumin (C) and piperine (P) after injection of $20 \mu \mathrm{~L}$ of the sample containing $10 \mu \mathrm{~g} / \mathrm{mL}$ C. longa and $5 \mu \mathrm{~g} / \mathrm{mL}$ P. nigrum extracts. Separation was carried out using a Knauer C18 column ( $250 \times 4.6 \mathrm{~mm}$, Eurospher 100 with $5 \mu \mathrm{~m}$ ) equipped with a pre-column and an isocratic mobile phase composed of an acetonitrile-methanol-water mixture with a volume ratio of 65:5:30 at flow rate of $1 \mathrm{~mL} / \mathrm{min}$ and a detection wavelength of 353 nm .
phase modifier [27]. Guan and Palmer studied the effect of a chaotropic agent trifluoroacetic acid on the retention of four weakly basic derivatives of triazole with different pKa values [28]. Using mixtures of water and acetonitrile as the mobile phase in the gradient mode, it was found that the lower the degree of protonation, the shorter the retention time. As phosphoric acid is also categorized as a chaotropic agent [29], it is presumable that the absence of piperine during 15 min running in this study using the mobile phase composition of acetonitrile-methanol-water-phosphoric acid $0.1 \%$ at a volume ratio of 20:30:47.5:2.5 ( pH 4.0 ) and acetonitrile-methanol-phosphoric acid $0.1 \%$ at 20:30:50 ( pH 3.0 ) might be attributed to the effect of chaotropic effect resulting partially protonated piperine, making piperine molecules interact with the stationary phase. However, when methanol was replaced by acetonitrile, a solvent with higher elution strength than methanol at the mobile phase composition of acetonitrile-water-phosphoric acid $0.1 \%$ of $50: 47.5: 2.5$ ( pH 4.0 ), the previous interaction of piperine with the stationary phase might be changed, in which piperine might be more solvated in the mobile phase. As a result, piperine was eluted at an early retention time of 0.516 min .

Based on the observation that the exchange of methanol by acetonitrile in the mobile phase and the presence of phosphoric acid had a dramatic effect on the retention time of piperine, various acetonitrile-methanol-water ratios without phosphoric acid were evaluated. In these experiments, phosphoric acid was not included in the mobile phase as acids can substantially reduce the life-time of the column. The effects of the mobile phase composition on peak retention (Tr), peak resolution (Rs), and peak tailing factor ( T ) are shown in Table 1.

When scrutinizing this table, it can be concluded that the optimal mobile phase composition consisted of an acetonitrile-methanol-water mixture with a volume ratio of 65:5:30, which corresponds to the calculated polarity index of 7.08 . Figure 4 shows the resulting baseline chromatographic separation of bis-demethoxycurcumin, demethoxycurcumin, curcumin, and piperine of the methanolic sample containing $10 \mu \mathrm{~g} / \mathrm{mL}$ C. longa and $5 \mu \mathrm{~g} / \mathrm{mL}$ P. nigrum extracts. Curcumin was eluted at 5.58 min with an Rs of 1.68 and a T of 1.16; piperine was eluted at 7.91
min with an Rs of 3.57 and T of 1.06. The obtained values of Rs, $\mathrm{T}, \mathrm{N}$ of curcumin and piperine meet the requirements of qualified peak in HPLC ( $\mathrm{T}<1.5$, Rs $>1.5$ and $\mathrm{N}>2000$ [17,29].

### 3.4. Specificity and selectivity

The method specificity is represented in Figure 5. There are no peaks observed at the retention time of curcumin and piperine in the blank sample (Figure 5a). The retention time for curcumin and piperine in the microparticle formulation sample (Figure 5c) is following the standard solution of curcumin and piperine (Figure 5b). The result indicates the method specificity. The method selectivity is also demonstrated by the baseline separation parameters obtained for curcumin and piperine at which the Rs values for curcumin and piperine are larger than 1.5 (Figure 5c). The curcumin peak was distinguishable from the demethoxycurcumin peak with the Rs of 1.6 and T of 1.1. These results confirm the selectivity of the developed method.

### 3.5. System suitability test

The optimized HPLC method was subjected to a system suitability test. A sample was injected six times in the system and the T, N, AUC, Tr, and Rs were determined. As shown in Table 2, the RSD values of all these parameters for curcumin as well as piperine were below $2 \%$ which indicates all parameters of the proposed HPLC method satisfy the USP and ICH standards. Therefore, the developed HPLC method is concluded to be suitable and effective for the analysis [17].

### 3.6. Linearity, range and sensitivity

Calibration curves were acquired by plotting peak area as a function of the corresponding concentration (Figure 6). Table 3 shows the regression parameters resulted from the least-square method. As shown in Figure 6 and Table 3, linearity was confirmed at concentration ranges


Figure 5. Chromatographic separation for specificity and selectivity assessment. (a) blank; (b) standard solution containing curcumin ( $10 \mu \mathrm{~g} / \mathrm{mL}$ ) and piperine ( $5 \mu \mathrm{~g} /$ mL ); (c) solid dispersion based microparticle ( $9.9 \mu \mathrm{~g} / \mathrm{mL}$ sample) containing C. longa and $P$. nigrum. $\mathrm{C}=$ curcumin, $\mathrm{P}=$ piperine, $\mathrm{BDMC}=$ bis-demethoxycurcumin, DMC $=$ demethoxycurcumin.

Table 2. System suitability test of the developed method.

| Injection | Tr (min) |  | AUC |  | T |  | Rs |  | N |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  | C | P | C | P | C | P | C | P | C | P |
| 1 | 5.421 | 7.660 | 226134 | 283307 | 1.205 | 1.065 | 1.624 | 3.574 | 8167.5 | 9561.8 |
| 2 | 5.588 | 7.914 | 229048 | 284215 | 1.238 | 1.071 | 1.649 | 3.576 | 8283.6 | 9356.3 |
| 3 | 5.642 | 7.970 | 230929 | 284790 | 1.215 | 1.072 | 1.680 | 3.578 | 8144.9 | 9243.9 |
| 4 | 5.652 | 7.994 | 231654 | 285454 | 1.205 | 1.098 | 1.690 | 3.576 | 8198.7 | 9409.2 |
| 5 | 5.644 | 7.974 | 231274 | 284831 | 1.197 | 1.097 | 1.688 | 3.577 | 8300.8 | 9474.6 |
| 6 | 5.636 | 7.977 | 231993 | 285380 | 1.202 | 1.094 | 1.690 | 3.575 | 8145.6 | 9353.2 |
| Mean | 5.597 | 7.915 | 230172 | 284662 | 1.210 | 1.083 | 1.670 | 3.576 | 8206.8 | 939987 |
| SD | 0.089 | 0.128 | 2230 | 803 | 0.015 | 0.015 | 0.027 | 0.001 | 69.2 | 109.8 |
| RSD (\%) | 1.595 | 1.614 | 0.969 | 0.282 | 1.221 | 1.389 | 1.646 | 0.040 | 0.84 | 1.17 |

$\mathrm{C}=$ Curcumin; $\mathrm{P}=$ Piperine; $\mathrm{Tr}=$ retention time; $\mathrm{Rs}=$ resolution; $\mathrm{T}=$ tailing factor; $\mathrm{N}=$ theoretical plate number; AUC $=$ Area Under Curve.


Figure 6. Calibration curves of piperine (a) and curcumin (b) standard following the method developed in this study ( $\mathrm{n}=3$ ). The series concentrations of calibration samples were $1.25 ; 2.50 ; 5.00 ; 7.50 ; 10.00 ; 12.50 ; 15.00 \mu \mathrm{~g} / \mathrm{mL}$ for piperine and $2.50 ; 5.00 ; 10.00 ; 15.00 ; 20.00 ; 25.00 ; 30.00 \mu \mathrm{~g} / \mathrm{mL}$ for curcumin.
of $1.25-15.00 \mu \mathrm{~g} / \mathrm{mL}$ and $2.00-30.00 \mu \mathrm{~g} / \mathrm{mL}$ for piperine and curcumin, respectively, as the correlation coefficient, r , within these ranges were higher than 0.999 . Also, linearity can be further evaluated by calculation of the RSD\% of the slope values, which did not exceed 2.0 \% [30].

Method sensitivity was tested by determining LOD and LOQ. The calculated LOD of piperine and curcumin was $0.27 \mu \mathrm{~g} / \mathrm{mL}$ and $0.42 \mu \mathrm{~g} /$ mL , respectively, and the calculated LOQ was $0.91 \mu \mathrm{~g} / \mathrm{mL}$ and $1.41 \mu \mathrm{~g} /$ mL for piperine and curcumin, respectively.

### 3.7. Accuracy and precision

Accuracy and precision studies were conducted according to the ICH recommendations [19] on three different levels of sample concentrations which are low, medium, and high of analyte concentration. The three concentration levels used in this study were $1.25 ; 7.50 ; 15.00$ and 2.50 ; $15.00 ; 30.00$ for piperine and curcumin, respectively.

Table 4 summarizes the accuracy and precision data. The results show the intra-day and inter-day recovery of piperine were between 99.54$101.50 \%$ and $99.38-99.89 \%$, respectively while the intra-day and interday recovery of curcumin reported were between $100.78-102.51 \%$ and 101.15-102.47\%, respectively. Repeatability and intermediate precision of the developed method were evaluated by calculating RSD obtained in one day and 3 different days on freshly prepared samples. As compiled in Table 4, it is shown that intra-day and inter-day RSD of piperine were $0.53-0.95 \%$ and $0.13-1.44 \%$, respectively; intra-day and inter-day RSD of curcumin were $0.28-1.621 \%$ and $0.46-1.14 \%$, respectively.

According to calculated recovery and RSD values, these reported data (Table 4) show that the recovery and RSD values fully satisfy within the generally accepted range of AOAC (90-107\% recovery and $<5.3 \%$ of

RSD) [21], which suggests a high level of accuracy and precision of the developed method.

### 3.8. Robustness

The robustness parameters of the developed method were obtained by investigating the impact of slight changes in the chromatographic conditions on peak retention time and peak area. Moderate changes in volume ratios of mobile phase composition ( $\pm 2 \%$ ), flow rate ( $\pm 0.1 \mathrm{~mL} /$ min ), and detection wavelength ( $\pm 2 \mathrm{~nm}$ ), however, did not significantly affect the retention time and peak area as indicated by the RSD value of $<2 \%$ (see Table 5).

### 3.9. Application of the developed method

The developed RP-HPLC method was applied to simultaneously determine the concentration of curcumin and piperine in the microparticle formulation sample containing multicomponent appears in C. longa and $P$. nigrum extracts. The developed method was successfully validated to separate multi-compounds in the microparticle formulation i.e. curcumin from its curcuminoids family, and piperine (Table 6). As shown in Table 6, the observed content of curcumin and piperine in the microparticles was $22.42 \pm 0.67 \mathrm{w} / \mathrm{w} \%$ and $9.04 \pm 0.67 \mathrm{w} / \mathrm{w} \%$, respectively. Using reference standards, it was found with the developed HPLC method that C. longa extract contained $75.38 \mathrm{w} / \mathrm{w} \%$ curcumin and $P$. nigrum extract contained $98.97 \mathrm{w} / \mathrm{w}$ \% piperine. As the claimed percentages of $C$. longa and $P$. nigrum extracts in the microparticles were $30 \mathrm{w} / \mathrm{w} \%$ and $10 \mathrm{w} / \mathrm{w} \%$, respectively, it can be calculated that the theoretical amounts of curcumin and piperine in the microparticles should be $22.61 \mathrm{w} / \mathrm{w} \%$

Table 3. Validation parameters for piperine and curcumin $(\mathrm{n}=3)$.

| Parameter | Piperine | Curcumin |
| :--- | :--- | :--- |
| Linearity range $(\mu \mathrm{g} / \mathrm{mL})$ | $1.25-15.00$ | $2.50-30.00$ |
| Linear equation | $\mathrm{Y}=140779 \mathrm{x}+11549$ | $\mathrm{Y}=42144 \mathrm{x}+1559.5$ |
| Correlation coefficient of $(\mathrm{r})$ | 0.9990 | 0.9994 |
| RSD of slope | 1.017 | 0.784 |
| Limit of Detection $(\mu \mathrm{g} / \mathrm{mL})$ | 0.27 | 0.42 |
| Limit of Quantification $(\mu \mathrm{g} / \mathrm{mL})$ | 0.91 | 1.41 |

between $90.0 \%$ and $110.0 \%$ [31]. Thus, the curcumin and piperine content found in the solid dispersion-based microparticle formulation using the developed method complies with the product specification of the USP monography. The results from the assay of curcumin and piperine confirm the successful implementation of the proposed HPLC method for the simultaneous quantification of curcumin and piperine in microparticles formulation containing multicomponent i.e. curcumin and piperine of $C$. longa and $P$. nigrum extracts. Furthermore, as the running time of HPLC analysis is only 10 min , the proposed method is suitable to

Table 4. Results of intra-day and inter-day accuracy and precision $\mathrm{n}=3$.

| Compound | Final concentration ( $\mu \mathrm{g} / \mathrm{ml}$ ) | Intra-day |  |  | Inter-day |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  |  | Recovered quantity ( $\mu \mathrm{g} / \mathrm{ml}$ ) | Recovery (\%) | RSD (\%) | Recovered quantity ( $\mu \mathrm{g} / \mathrm{ml}$ ) | Recovery (\%) | RSD (\%) |
| Curcumin | 2.50 | $2.54 \pm 0.04$ | 101.49 | 1.47 | $2.61 \pm 0.02$ | 102.47 | 0.79 |
|  | 15.00 | $15.12 \pm 0.25$ | 100.78 | 1.62 | $15.28 \pm 0.17$ | 101.88 | 1.14 |
|  | 30.00 | $30.75 \pm 0.09$ | 102.51 | 0.28 | $31.39 \pm 0.14$ | 101.15 | 0.46 |
| Piperine | 1.25 | $1.27 \pm 0.01$ | 101.50 | 0.95 | $1.25 \pm 0.01$ | 99.79 | 1.08 |
|  | 7.50 | $7.47 \pm 0.07$ | 99.54 | 0.94 | $7.45 \pm 0.01$ | 99.38 | 0.13 |
|  | 15.00 | $15.21 \pm 0.08$ | 101.38 | 0.53 | $14.98 \pm 0.22$ | 99.89 | 1.44 |

Table 5. Robustness test of the developed method $(\mathrm{n}=3)$.

| Condition | Curcumin |  |  |  | Piperine |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  | AUC |  | $\mathrm{Tr}(\mathrm{min})$ |  | AUC |  | Tr (min) |  |
|  | mean | RSD (\%) | mean | RSD (\%) | mean | RSD (\%) | mean | RSD (\%) |
| Selected mobile phase composition is acetonitrile: methanol: water $=65: 5: 30 \mathrm{vol}-\%$ |  |  |  |  |  |  |  |  |
| 60:10:30 vol-\% | 21273.6 | 0.72 | 5.58 | 0.10 | 25209.7 | 0.308 | 7.84 | 0.09 |
| 70:5:25 vol-\% | 22796.7 | 0.54 | 5.66 | 0.10 | 25358.3 | 0.500 | 7.95 | 0.49 |
| Flow rate of $1 \mathrm{~mL} / \mathrm{min}$ |  |  |  |  |  |  |  |  |
| $0.9 \mathrm{~mL} / \mathrm{min}$ | 21320.7 | 0.32 | 5.49 | 0.02 | 25296.3 | 0.955 | 7.83 | 0.05 |
| $1.1 \mathrm{~mL} / \mathrm{min}$ | 21255.7 | 0.83 | 5.82 | 0.43 | 25164.6 | 0.501 | 7.99 | 0.12 |
| Selected detection wavelength of 353 nm |  |  |  |  |  |  |  |  |
| 351 nm | 21120.3 | 1.15 | 5.61 | 0.16 | 325481.6 | 0.329 | 7.85 | 0.63 |
| 355 nm | 23781.4 | 0.45 | 5.63 | 0.07 | 252137.6 | 0.264 | 7.92 | 0.17 |

$\underline{\mathrm{Tr}}=$ retention time; AUC $=$ Area Under Curve.
and $9.90 \mathrm{w} / \mathrm{w} \%$, respectively. Using these percentages, it can be calculated that the recovery of curcumin and piperine from the microparticles was $99.14 \%$ and $91.31 \%$, respectively, using the developed HPLC method (Table 6).

USP monograph on curcuminoids dosage form requires that the content of the active compound in the dosage form products should be

## Table 6. Curcumin and piperine content in microparticles.

| Sample | Amount found in microparticles |  |
| :--- | :--- | :--- |
|  | Curcumin (w/w \%) | Piperine (w/w \%) |
| 1 | 21.17 | 8.41 |
| 2 | 22.90 | 8.39 |
| 3 | 22.58 | 8.55 |
| 4 | 22.90 | 9.98 |
| 5 | 22.82 | 9.40 |
| 6 | 22.17 | 9.50 |
| Mean | 22.42 | 9.04 |
| SD | 0.67 | 0.67 |
| RSD $(\%)$ | 3.05 | 7.47 |

Formulation claim: $30 \mathrm{w} / \mathrm{w} \%$ C. longa and $10 \mathrm{w} / \mathrm{w} \%$ P. nigrum extracts.
be applied during routine analysis as part of quality control procedures in the formulation of the microparticles.

## 4. Conclusion

A suitable, rapid, accurate, and precise RP-HPLC method was developed for the simultaneous determination of curcumin and piperine in a solid dispersion-based microparticle formulation. The developed method offers a linear response across a wide range of analyte concentrations with satisfactory method sensitivity. Furthermore, the proposed RP-HPLC method guarantees the extension of the column lifetime and HPLC system due to the absence of acid in the mobile phase and the use of commonly used organic solvents, i.e. acetonitrile and methanol.

## Declarations

## Author contribution statement

Dewi Setyaningsih: Conceived and designed the experiments; Analyzed and interpreted the data; Contributed reagents, materials, analysis tools or data; Wrote the paper.

Yosua Agung Santoso: Performed the experiments.
Yustina Sri Hartini: Analyzed and interpreted the data.

Yosi Bayu Murti: Contributed reagents, materials, analysis tools or data.

Wouter L. J. Hinrichs: Conceived and designed the experiments; Analyzed and interpreted the data; Wrote the paper.

Christine Patramurti: Conceived and designed the experiments; Performed the experiments.

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## Data availability statement

Data included in article/supplementary material/referenced in article.

## Declaration of interests statement

The authors declare no conflict of interest.

## Additional information

No additional information is available for this paper.

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